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(54) **PREPARATION SYSTEM AND METHOD**

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This patent is subject to a terminal disclaimer.

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(51) **Int. Cl.**
G01N 21/75 (2006.01)
C07H 21/00 (2006.01)
C12M 1/00 (2006.01)

(52) **U.S. Cl.**
CPC **C12M 45/09** (2013.01); **C07H 21/00** (2013.01); **G01N 21/75** (2013.01)

(58) **Field of Classification Search**

CPC C07H 21/00; G01N 21/75

USPC 536/25.4; 422/417

See application file for complete search history.

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(57) **ABSTRACT**

Systems and methods for preparing a sample for further analysis are provided. The system can include an enclosure. A membrane can be disposed within the enclosure. First and second reservoirs can be disposed within the enclosure, and at least one of the first and second reservoirs can be adapted to have a reagent disposed therein. A valve can be disposed within the enclosure and in fluid communication with the first or second reservoirs or both. The valve can also be in fluid communication with the membrane. The valve can be adapted to selectively regulate the flow of the reagent from the first reservoir, through the membrane, and into the second reservoir.

17 Claims, 10 Drawing Sheets

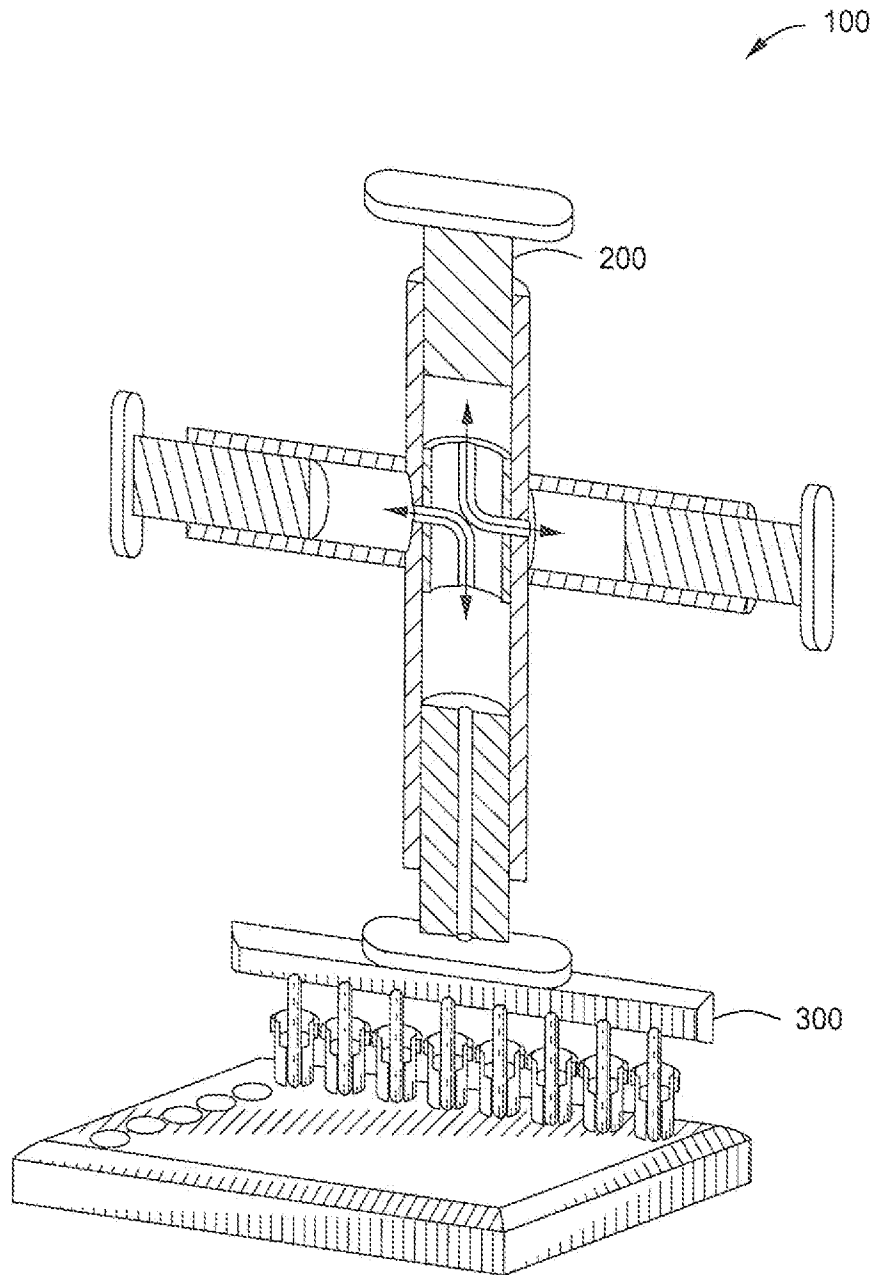


FIG. 1

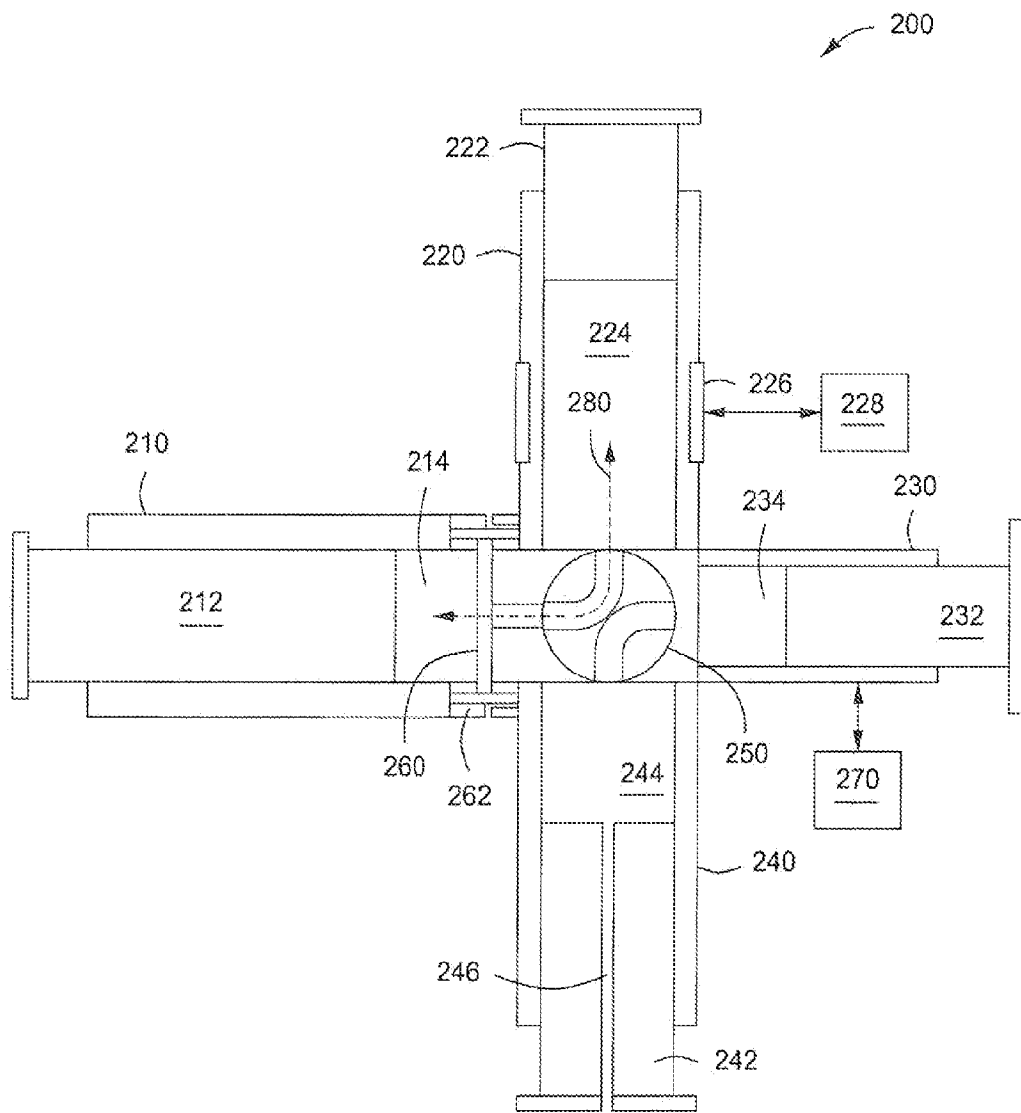


FIG. 2A

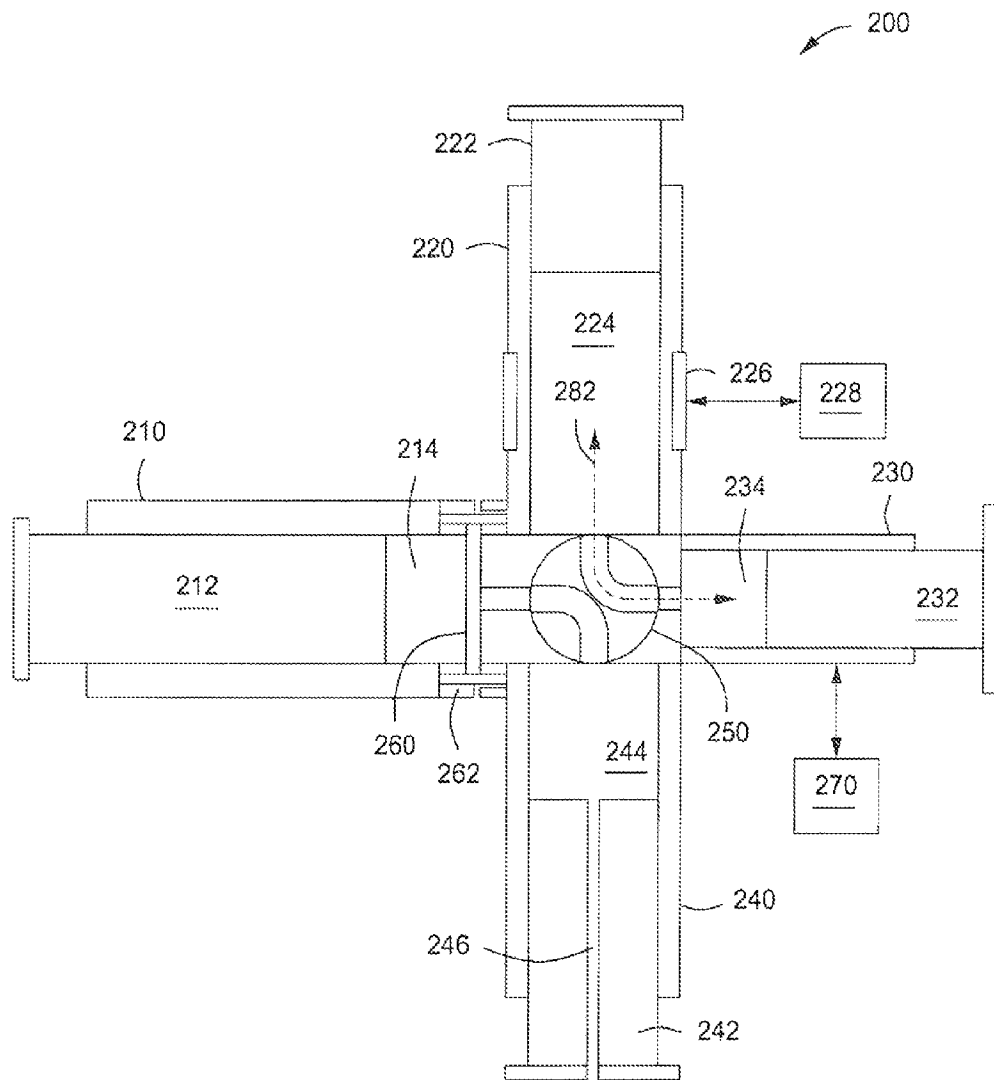


FIG. 2B

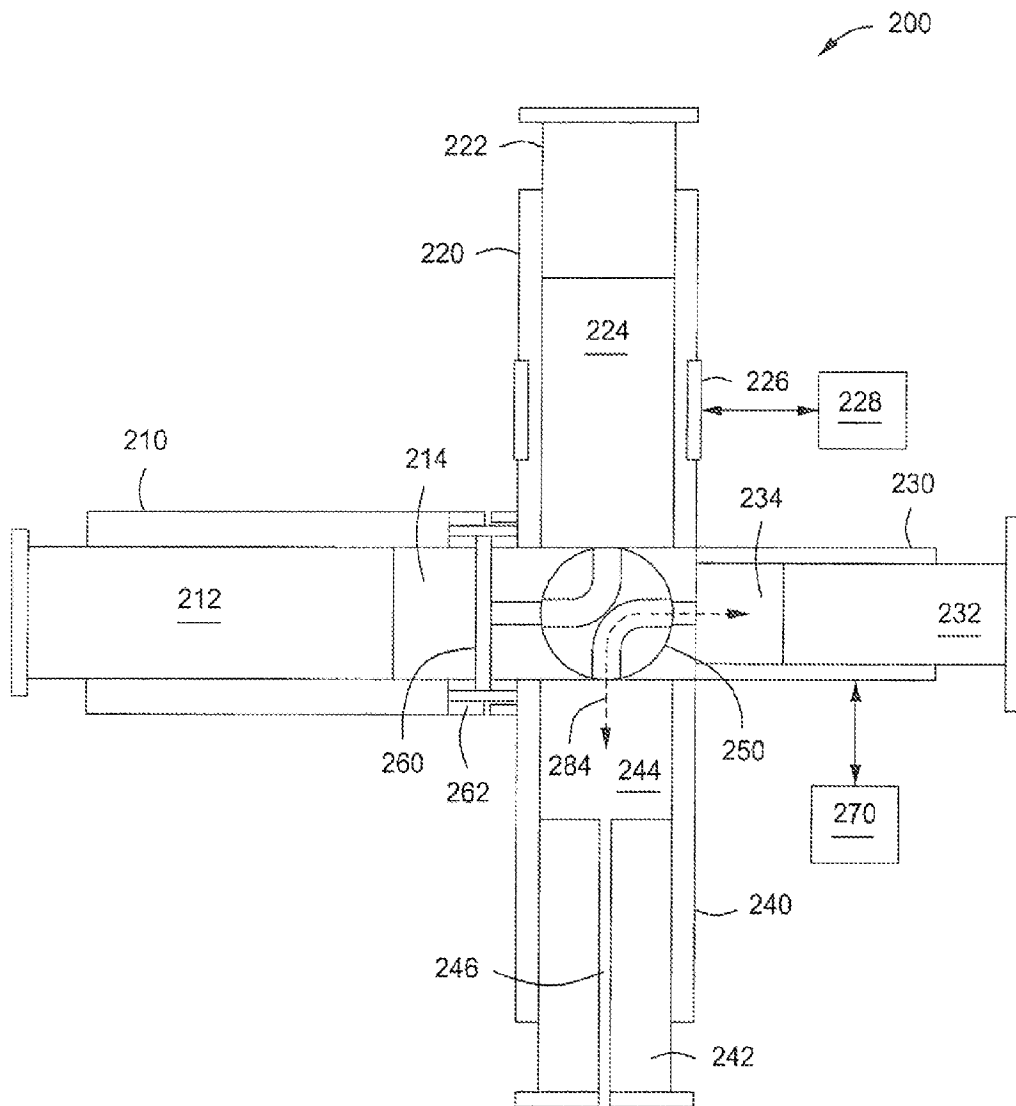


FIG. 2C

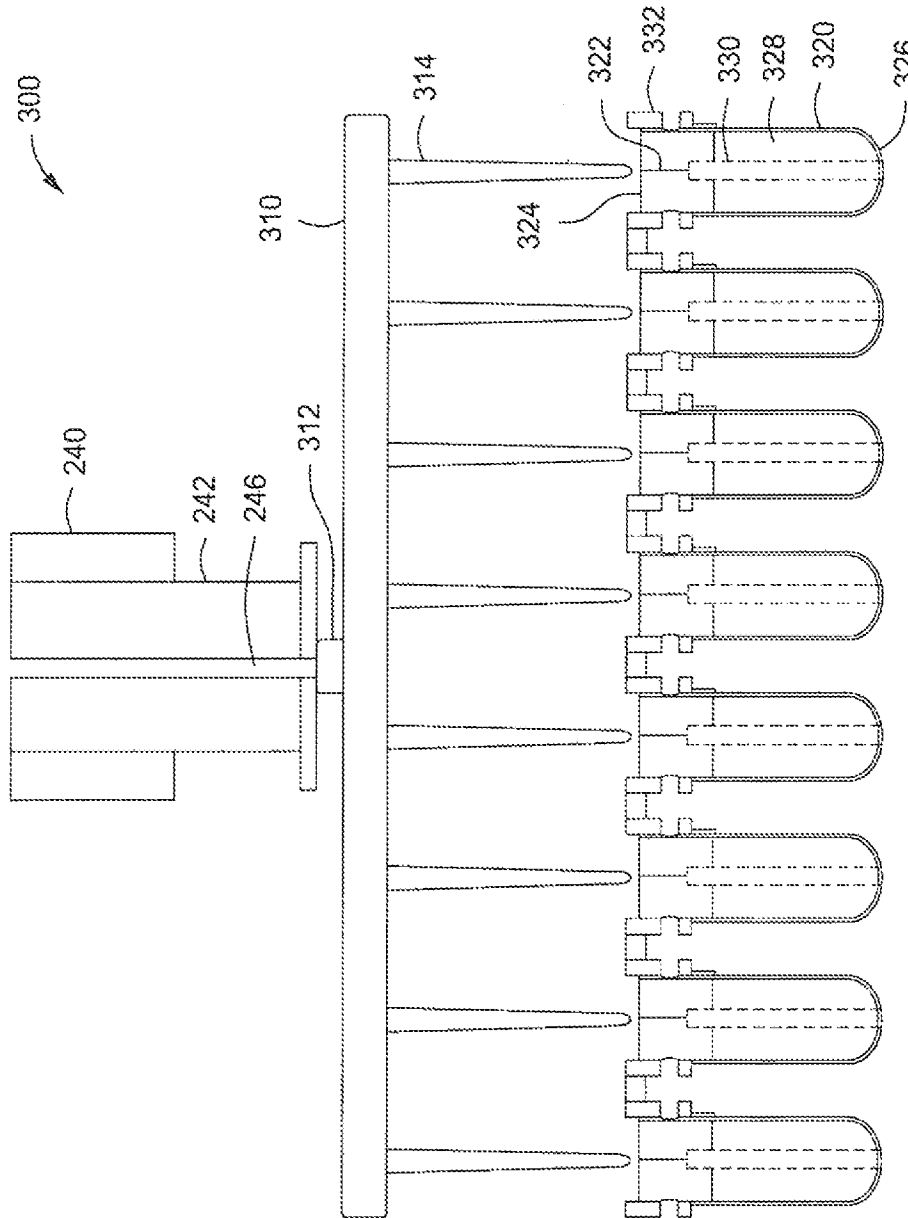


FIG. 3

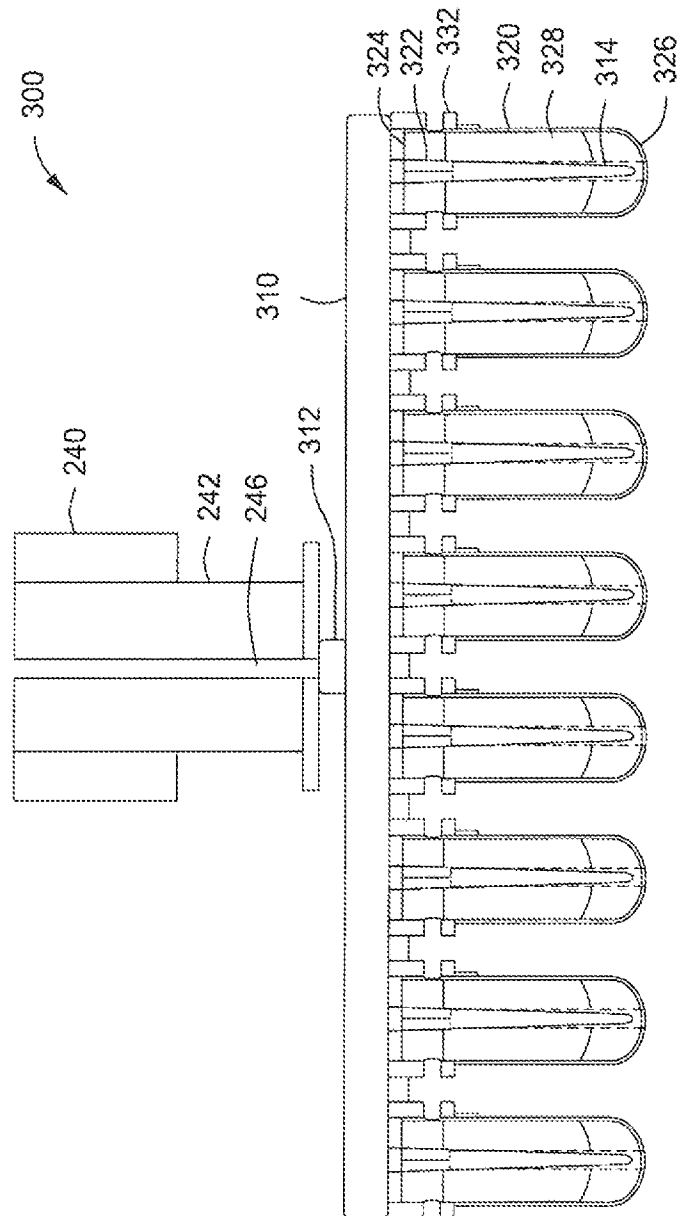


FIG. 4

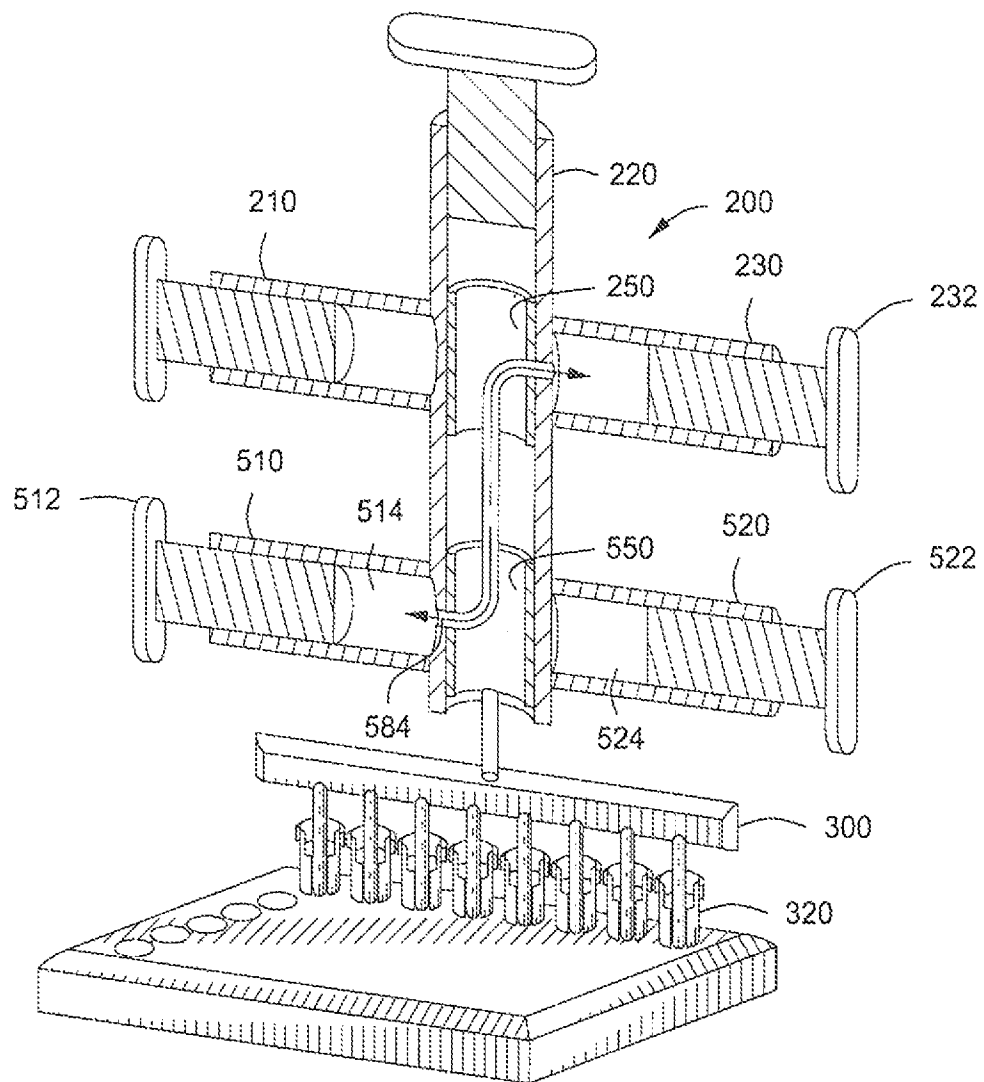


FIG. 5A

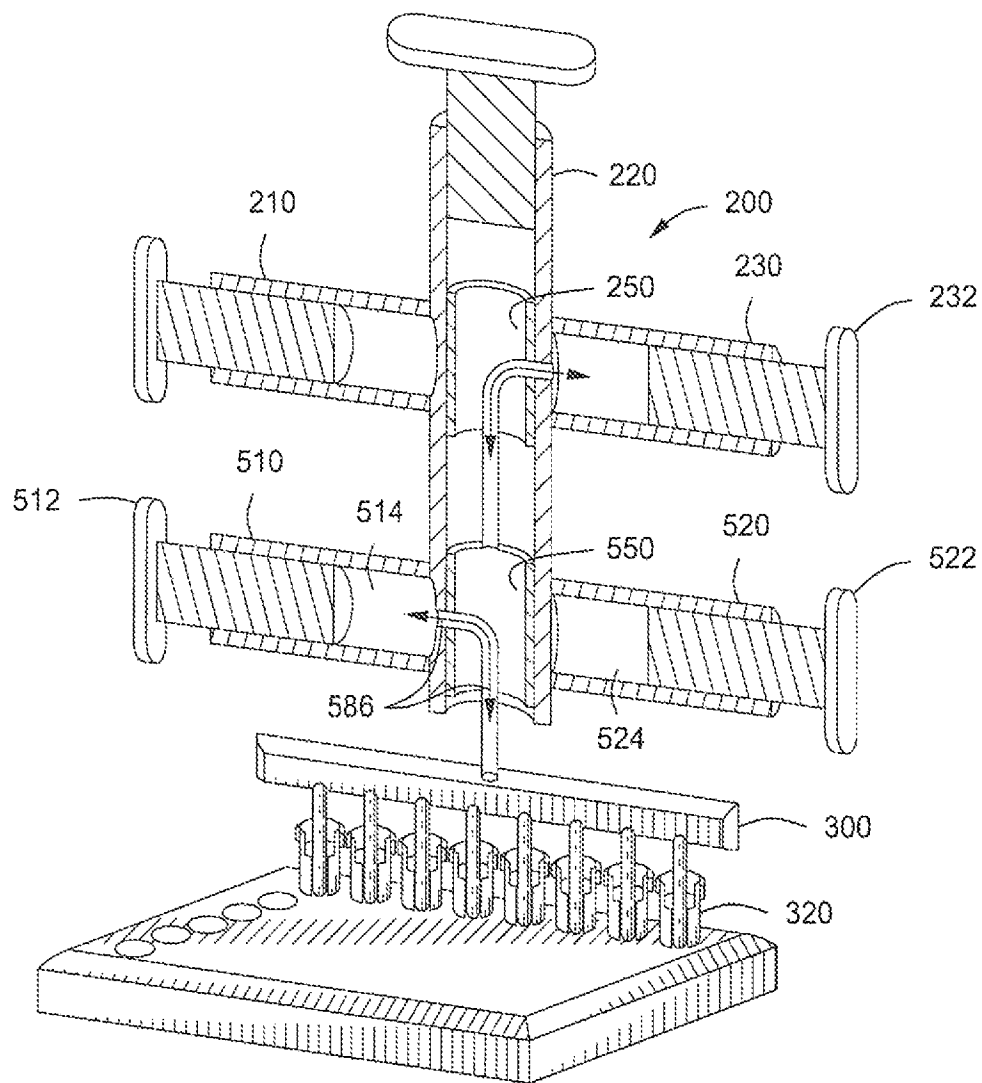


FIG. 5B

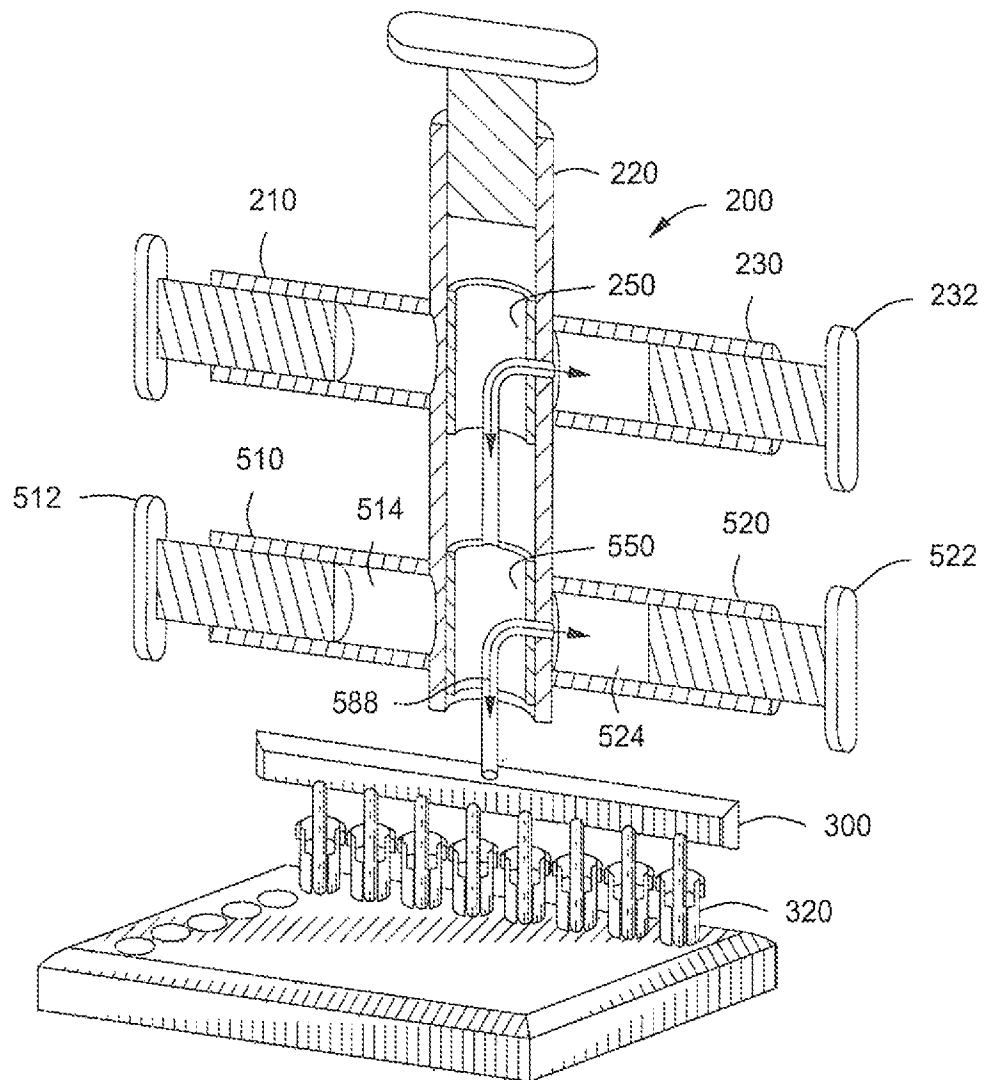


FIG. 5C

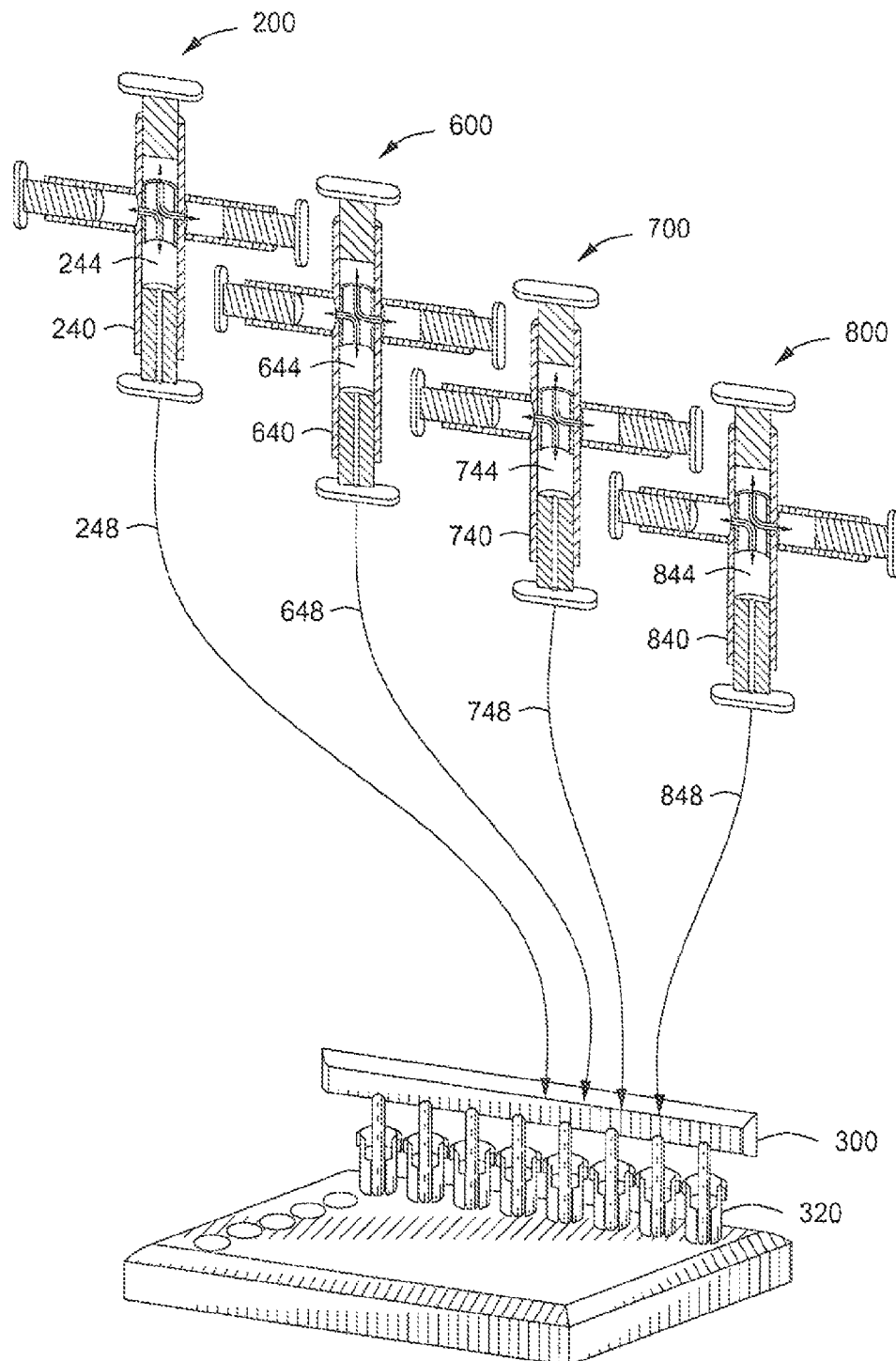


FIG. 6

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PREPARATION SYSTEM AND METHOD**CROSS-REFERENCE TO RELATED APPLICATIONS**

This application claims the benefit of a related U.S. Provisional Application Ser. No. 61/680,075 filed Aug. 6, 2012, entitled "Pre-Polymerase Chain Reaction Preparation Kit," the disclosure of which is incorporated by reference herein in its entirety.

ORIGIN OF THE INVENTION

The invention described herein was made in the performance of work under a NASA contract and is subject to the provisions of Section 305 of the National Aeronautics and Space Act of 1958, Public Law 85-568 (6052 Stat. 435; 42 U.S.C. 233605).

BACKGROUND**Field**

Embodiments described herein generally relate to apparatus, systems, and methods for the isolation and/or preparation of biological and non-biological samples for further analysis, such as pre-analysis sample-reagent mixing, distributing, and loading.

SUMMARY

This summary is provided to introduce a selection of concepts that are further described below in the detailed description. This summary is not intended to identify key or essential features of the claimed subject matter, nor is it intended to be used as an aid in limiting the scope of the claimed subject matter.

Systems and methods for releasing biological and non-biological samples of interest, such as nucleic acids or other samples, from a membrane are disclosed. The system, which may be self-enclosed and automated for use in space or terrestrial environments, can include an enclosure. A membrane can be disposed within the enclosure. First and second reservoirs can be disposed within the enclosure, and at least one of the first and second reservoirs can have a reagent disposed therein. A valve, such as a multi-way valve, can be disposed within the enclosure and in fluid communication with the first or second reservoirs, or both. The valve can also be in fluid communication with the membrane. The valve can be adapted to selectively regulate the flow of the reagent from the first reservoir, through the membrane, and into the second reservoir. The system can be used for sample-reagent mixing, distributing, and loading. In one embodiment, when the system is used, an analyte or other sample can be isolated or prepared for further analysis, such as real-time polymerase chain reaction, without exposure to its external environment.

The method can include flowing a first reagent with liquid samples through a first flow path between a first reservoir and a second reservoir. The first flow path can extend through a valve, or a valve and a membrane. An analyte of interest, such as nucleic acids or other samples, can be released from the membrane when contacted with the first reagent to form a first mixture including the first reagent and the released nucleic acids or other samples. Or, liquid samples can be directly introduced to the second reservoir from the first reservoir through the valve. The valve can be rotated to form a second flow path between the second reservoir and a third reservoir

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through the valve. At least a portion of the first mixture can flow through the second flow path and into the third reservoir containing a second reagent to form a second mixture. The method can be performed to enable elution, dilution, mixing, distribution, or loading of a sample.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 depicts a cross-sectional perspective view of an illustrative sample preparation system, according to one or more embodiments described.

FIGS. 2A-2C depict cross-sectional views of the preparation assembly depicted in FIG. 1 during a process for preparation of a sample, according to one or more embodiments described.

FIG. 3 depicts a cross-sectional view of the fourth injector of the preparation assembly in fluid communication with a distributor, according to one or more embodiments described.

FIG. 4 depicts a cross-sectional view of the distributor in fluid communication with one or more containers, according to one or more embodiments described.

FIGS. 5A-5C depict partial cross-sectional perspective views of another illustrative embodiment of a sample preparation assembly having a second valve in fluid communication with the containers via the distributor, according to one or more embodiments described.

FIG. 6 depicts a perspective view of a plurality of preparation assemblies in fluid communication with the containers via the distributor, according to one or more embodiments described.

DETAILED DESCRIPTION

FIG. 1 depicts a cross-sectional perspective view of an illustrative sample preparation system **100**, according to one or more embodiments described herein. The sample preparation system **100** can include one or more elution/sample preparation assemblies **200** and one or more distribution assemblies **300**. The sample preparation system **100** is a pipette-free, closed system adapted to limit or prevent exposure to an external environment. In one embodiment, the sample preparation system **100** can be used in a microgravity environment. As a closed system, fluid disposed within the system **100** does not escape or leak out, as would occur with traditional open-ended pipettes, particularly when used in microgravity environments in space.

FIGS. 2A-2C depict cross-sectional views of the preparation assembly **200** depicted in FIG. 1 during a process for preparation of a sample, according to one or more embodiments. The preparation assembly **200** can include one or more injectors (four are shown **210**, **220**, **230**, **240**), one or more valves (one is shown **250**), and one or more membranes (one is shown **260**). Although not shown, in at least one embodiment, the preparation assembly **200** can include two injectors **210**, **220** having the membrane **260** disposed therebetween. In this embodiment, the additional injectors **230**, **240** and the valve **250** can be omitted.

The injectors **210**, **220**, **230**, **240** can include one or more components. For example, each injector **210**, **220**, **230**, **240** can have a chamber or reservoir **214**, **224**, **234**, **244** disposed therein, respectively. Each injector **210**, **220**, **230**, **240** can also have a piston **212**, **222**, **232**, **242**, respectively, adapted to slide axially therein. Axial movement of the piston **212**, **222**, **232**, **242** within its respective injector **210**, **220**, **230**, **240** can push fluid out of its respective reservoir **214**, **224**, **234**, **244** and into one of the reservoirs **214**, **224**, **234**, **244** of another injector. Similarly, axial movement of the piston **212**, **222**,

232, 242 within its respective injector **210, 220, 230, 240** can also draw fluid into its respective reservoir **214, 224, 234, 244** from one of the reservoirs **214, 224, 234, 244** of another injector.

At least one of the pistons **212, 222, 232, 242** can have a bore formed axially therethrough such that the bore forms a path of fluid communication between the reservoir and the exterior of the injector **210, 220, 230, 240**. As shown in FIG. 2A, the fourth piston **242** has a bore **246** formed therethrough. The bore **246** can allow fluid to be injected into or withdrawn from the reservoir **244** of the fourth injector **240**, as described in more detail below.

At least one of the injectors **210, 220, 230, 240** can have an interface **226** adapted to receive a light source **228**, such as a spectrophotometer, illuminometer, and/or fluorometer. The light source **228** can be adapted to measure the reflection, emission, or transmission properties of the fluid within the reservoir **224** as a function of its wavelength. The interface **226** can be any shape, including round, flat, or the like that fits the adapted light source **228**. In at least one embodiment, the light source **228** can be a nanodrop (ND) 2000c spectrophotometer.

The valve **250** can be disposed within the preparation assembly **200** and coupled to at least one of the injectors **210, 220, 230, 240** such that the reservoirs **214, 224, 234, 244** of the injectors **210, 220, 230, 240** can be in fluid communication with one another through the valve **250**. In at least one embodiment, one or more of the injectors **210, 220, 230, 240** can be operatively connected, such as by threadable engagement or other method of attachment, with the valve **250**. Although not shown, tubing having a diameter similar to the diameter of the flow path through the valve **250**, e.g., 15-gauge, can be coupled to and disposed between the injectors **210, 220, 230, 240** and the valve **250** to provide more flexibility in the layout or configuration of the preparation assembly **200**.

The valve **250** can include a rotatable housing having at least one bore formed therethrough. In FIGS. 1 and 2A-2C, the valve **250** has two bores shown, but one bore will be discussed in further detail with reference to the FIGS. 2A-2C during operation of the preparation system. The valve **250** can be actuated or rotated to provide more than one path of fluid communication among the plurality of reservoirs **214, 224, 234, 244**, i.e., the valve may be considered to be a "multi-way" valve. In one embodiment, a suitable valve **250** can be or include those manufactured by the Hamilton Company in Reno, Nev.

In at least one embodiment, the membrane **260** can have isolated nucleic acids, such as DNA and/or RNA disposed thereon. The term "membrane," as used herein, refers to any material or layer(s) of material that acts as a selective barrier, allowing fluids and some particles to pass therethrough, while other particles are not allowed to pass therethrough, and are instead retained therein.

The membrane **260** can be disposed within a membrane support assembly **262**, and the membrane support assembly **262** can be disposed within the preparation assembly **200**. In at least one embodiment, the membrane support assembly **262** can be disposed between one of the injectors **210, 220, 230, 240** and the valve **250**. For example, the membrane support assembly **262** can be operatively connected, such as by being threadably engaged or otherwise attached, with the first injector **210** and the valve **250**. The support assembly **262** can be perforated and disposed adjacent to opposing surfaces of the membrane **260**. As such, fluid can pass or flow through the membrane **260** and the membrane support assembly **262**.

A suitable membrane **260** can include one or more micropores. In one embodiment, the micropores can have an average pore diameter ranging from a low of about 0.2 μm , about 0.4 μm , or about 0.6 μm to a high of about 1 μm , about 2 μm , about 3 μm , about 4 μm , about 5 μm , or more for RNA/DNA isolation and/or preparation. However, for other applications, the micropores can have an average pore diameter ranging from a low of about 0.0001 μm , about 0.001 μm , or about 0.01 μm to a high of about 0.05 μm , about 0.1 μm , or about 0.2 μm (e.g., for protein isolation or separation), or from a low of about 10 μm , about 20 μm , or about 30 μm to a high of about 50 μm , about 75 μm , or about 100 μm (e.g., for filtering single cell suspensions). The membranes can also be used for binding, isolating, and separating specific molecules and particles, isolating specific types of cells, and reagent sterilization.

The membrane **260** can be, but is not limited to, a proteinase membrane, a homogenizing membrane, a filtering membrane, a binding membrane, or any combination thereof. For example, the membrane **260** can be a binding membrane having precipitated nucleic acids bound thereto. Suitable membranes **260** can also include a solid surface functionalized with immobilized active enzymes and high density enzyme surfaces. More specifically, suitable membranes **260** can include a solid surface functionalized with immobilized proteinase, such as trypsin, chymotrypsin, endoproteinase GluC, papain, endoproteinase pepsin, proteinase K, and the like. Suitable solid surfaces can also include glass fiber, glass fiber treated with oleophobic coatings, silica particles or beads, silica particles or beads coated with oleophobic coatings, nylon, and other oleophobic materials, and the like. For example, suitable membranes can include the DigesTip produced by ProteoGen Bio in Siena, Italy. Suitable membranes **260** can also include, but are not limited to, membranes found in a standard kit for the purification of DNA or RNA, such as the commercially available kit sold by the manufacturer Qiagen, (e.g., DNeasy Blood and Tissue Kit, RNeasy Mini Kit, RNeasy Protect Mini Kit, and RNeasy Plant Mini Kit). In at least one embodiment, the surface can be coated with complementary oligo nucleotides, antibodies, covalently or non-covalently binding to target molecules, and/or any particles and membranes used for separating molecules based on size and isoelectric point.

One or more control systems **270** can be in communication with the preparation assembly **200**. In at least one embodiment, the control system **270** can be adapted to measure the volume of fluid within the reservoirs **214, 224, 234, 244**, measure the concentration of nucleic acids disposed in the fluid within the reservoirs **214, 224, 234, 244**, and/or actuate the pistons **212, 222, 232, 242** to cause a calculated amount of fluid to flow between reservoirs **214, 224, 234, 244**. The control system **270** can serve as a digital volume control and automatic distributor of fluid within the preparation assembly **200**.

FIG. 3 depicts a cross-sectional view of the fourth injector **240** of the preparation assembly **200** in fluid communication with a distributor or manifold **310** of distribution assembly **300**, and FIG. 4 depicts a cross-sectional view of the distributor **310** in fluid communication with one or more containers or tubes **320**, according to one or more embodiments. The distribution assembly **300** can include the distributor **310** and one or more tubes or containers (eight are shown **320**). The distributor **310** can include an inlet **312** in fluid communication with one or more outlets **314**. The outlets **314** can be made of any material (plastic, metal, etc.) and sized to fit within the containers **320**. In at least one embodiment, the inlet **312** can be coupled to the fourth injector **240** directly, or

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through a flexible conduit or tube to allow more adaptability in configuration and to allow multi-sample preparation. Further, the inlet **312** may be configured to be in fluid communication with the fourth reservoir **244** through the bore **246** in the piston **242**. Each outlet **314** can be adapted to be inserted into a container **320**, as shown in FIG. 4. The number of outlets **314** and/or containers **320** can range from a low of 1, 2, 4, 6, 8, or 10 to a high of about 15, 20, 25, 30, 35, 40, or more. The distributor **310** can be designed with additional inlets to communicate with the exterior to allow washing and creation of a vacuum within each container **320**.

The containers **320** can be made of glass or plastic and define an internal volume. The internal volume can range from a low of about 0.25 mL, about 0.5 mL, about 1 mL, about 2 mL, about 3 mL, about 4 mL, or about 5 mL to a high of about 6 mL, about 8 mL, about 10 mL, about 15 mL, about 20 mL, or more. Each container **320** can include a seal **322** (e.g., a rubber seal) proximate a first end **324** thereof. The seal **322** can be adapted to prevent fluid from leaking into or out of the container **320**. Each container **320** can also have a piston or plug **328** disposed therein and coupled to the seal **322**. The plug **328** can be adapted to sealingly engage the inner walls of the container **320** and slide axially therein. The plug **328** can have a bore formed axially therethrough, and a tube **330** can be disposed within the bore. The tube **330** can have a diameter ranging from a low of about zero (0) (e.g., when the outlet **314** is not inserted), about 0.5 mm, about 1 mm, or about 2 mm to a high of about 4 mm, about 6 mm, about 8 mm, or more. Each tube **330** can be adapted to have one of the outlets **314** of the distributor **310** inserted into it.

Accordingly, each outlet **314** can extend through the seal **322**, the plug **328**, and/or the tube **330** to introduce fluids into the corresponding container **320**. Once the outlet **314** is inserted into and in fluid communication with the container **320**, the plug **328** can be moved toward the first end **324** of the container **320** generating a vacuumed space between the plug **328** and a second end **326** of the container **320**. At the same time, the piston **242** (in this example the representative injector connected is injector **240**) will move up to release the mixture from the fourth reservoir **244**. For example, an actuator **332** can be coupled to the container **320** and adapted to move or pull the corresponding plug **328** toward the first end **324** of the container **320** to create the vacuumed space. In at least one embodiment, each container **320** can include its own actuator **332**, and the actuators **332** can be coupled together to move a plurality of plugs **328** simultaneously. In another embodiment, a single actuator **332** can be adapted to move the plurality of plugs **328** in a plurality of containers **320** simultaneously. The vacuumed space can induce the fluid from the fourth reservoir **244** to flow through the bore **246** in the piston **242** and the distributor **310** and into one or more of the containers **320**, as described in more detail below. Once the fluid is introduced, the seal **322** can prevent the fluids from leaking into or out of the container **320**.

Now referring to FIGS. 1-4, in operation, the membrane **260** having the sample of interest, such as nucleic acids, bound thereto can be disposed within the membrane support assembly **262**, and the membrane support assembly **262** can be operatively positioned within the preparation assembly **200**. For example, the membrane support assembly **262** can be threadably engaged with the first injector **210** and the valve **250**.

The valve **250** can be rotated to provide a first flow path **280** between the first and second reservoirs **214**, **224** (see FIG. 2A). The first flow path **280** can extend through the valve **250** and the membrane **260**. The second reservoir **224** can include a first reagent. The volume of the first reagent in the second

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reservoir **224** can range from a low of about 0.1 mL, about 0.2 mL, about 0.4 mL, about 0.6 mL, about 0.8 mL, or about 1.0 mL to a high of about 2.0 mL, about 3.0 mL, about 4.0 mL, about 5.0 mL, or more. In at least one embodiment, the first reagent can include water. In another embodiment, the first reagent can be a TE buffer (such as 10 mM Tris-Cl/1 mM EDTA, having a pH of about 7.5), an EB buffer (such as 10 mM Tris-Cl, having a pH of about 8.5), or the like. The second piston **222** can be pressed, thereby forcing the first reagent through the first flow path **280** and into the first reservoir **214**. As the first reagent contacts the first membrane **260**, the first reagent can cause the sample of interest, such as nucleic acids (e.g., DNA and/or RNA), bound to the first membrane **260** to release therefrom and become disposed or suspended in the first reagent, forming a first mixture in the first reservoir **214**.

The first piston **212** can then be pressed, thereby forcing the first mixture back through the first flow path **280** and into the second reservoir **224**. The concentration of the samples, such as the nucleic acids, within the first mixture can be measured. For example, the control system **270**, operatively connected and using the light source **228**, can measure the concentration of the samples of interest, such as nucleic acids, within the first mixture.

Referring now to FIG. 2B, the valve **250** can then be rotated to provide a second flow path **282** between the second and third reservoirs **224**, **234**. The third reservoir **234** can include a second reagent. The volume of the second reagent in the third reservoir **234** can range from a low of about 0.1 mL, about 0.2 mL, about 0.4 mL, about 0.6 mL, about 0.8 mL, or about 1.0 mL to a high of about 2.0 mL, about 3.0 mL, about 4.0 mL, about 5.0 mL, or more. In at least one embodiment, the second reagent can include water or any other suitable dilution buffer. The second piston **222** can be pressed, thereby forcing at least a portion the first mixture through the second flow path **282** and into the third reservoir **234**, forming a second mixture including the first mixture and the second reagent in the third reservoir **234**.

The amount of the first mixture introduced to the third reservoir **234** can depend, at least in part, on the concentration of the samples of interest, such as the concentration of nucleic acids disposed within the first mixture. For example, as the amount of the first mixture introduced to the third reservoir **234** increases, the concentration of the second mixture increases; however, as the amount of the first mixture introduced to the third reservoir **234** decreases, the second mixture becomes more diluted. In at least one embodiment, a ratio of DNA to the total volume of the first and/or second mixture can range from a low of about 0.4 ng/ μ L, about 0.8 ng/ μ L, about 1.2 ng/ μ L, about 1.6 ng/ μ L, or about 2.0 ng/ μ L to a high of about 3.0 ng/ μ L, about 4.0 ng/ μ L, or about 5.0 ng/ μ L. In another embodiment, a ratio of RNA to the total volume of the first and/or second mixture can range from a low of about 0.05 ng/ μ L, about 0.10 ng/ μ L, about 0.20 ng/ μ L, about 0.30 ng/ μ L, or about 0.40 ng/ μ L to a high of about 1.0 ng/ μ L, 1.2 ng/ μ L, about 1.4 ng/ μ L, about 1.6 ng/ μ L, about 1.8 ng/ μ L, or about 2.0 ng/ μ L.

The valve **250** can then be rotated to provide a third flow path **284** between the third and fourth reservoirs **234**, **244** (see FIG. 2C). The fourth reservoir **244** can include a third reagent. The volume of the third reagent in the fourth reservoir **244** can range from a low of about 0.1 mL, about 0.2 mL, about 0.4 mL, about 0.6 mL, about 0.8 mL, or about 1.0 mL to a high of about 2.0 mL, about 3.0 mL, about 4.0 mL, about 5.0 mL, or more. In at least one embodiment, the third reagent can include a Polymerase Chain Reaction (PCR) master mix. The PCR master mix can include DNA polymerase, a buffer, and deoxynucleoside triphosphates (dNTPs). Suitable PCR mas-

ter mixes can include the Taq PCR Master Mix commercially available from Qiagen, Inc. in Valencia, Calif. In at least one embodiment, when the second mixture includes RNA as the sample of interest, the second mixture can be mixed with reverse transcriptase prior to being combined with the PCR master mix in the fourth reservoir **244**. In another embodiment, the reverse transcription polymerase chain reaction can be completed in a single step. When the preparation assembly **200** is coupled with another assembly (not shown), such as the isolation assembly shown and described in U.S. patent application Ser. No. 13/461,487 entitled "System and Method for Isolation of Samples," the entire content of which is hereby incorporated herein by reference, the RNA and reverse transcriptase can be injected first to complete reverse transcription, and then PCR mix can be injected from another reservoir to complete PCR.

Following the rotation of valve **250** to provide third flow path **284**, the third piston **232** can be pressed, thereby forcing at least a portion of the second mixture through the third flow path **284** and into the fourth reservoir **244**, forming a third mixture including the second mixture and the third reagent. In at least one embodiment, the valve **250** can then be rotated to form the flow path **280** (as shown in FIG. 2A) or the flow path **282** (as shown in FIG. 2B), particularly when only one bore is present within valve **250**, such that the valve **250** is "closed" to the fourth reservoir **244**. In another embodiment, a second bore may be present within valve **250** that is constructed with a check valve to permit flow of fluid in only one direction, such as in this embodiment from the third reservoir **234** to the fourth reservoir **244**, thereby preventing back flow of the third mixture and effectively "closing" the fourth reservoir **244** in an alternate manner. In at least one embodiment, the actuators **332** can then move the plugs **328** toward the first end **324** of the containers **320** inducing the vacuumed space between the plugs **328** and the second end **326** of the containers **320** (see FIGS. 3 and 4). The vacuumed space can induce the third mixture to flow from the fourth reservoir **244** through the bore **246** in the piston **242** and toward the distributor **310**. In another embodiment, the piston **242** of the fourth injector **240** can be pressed forcing the third mixture through the bore **246** in the piston **242** and toward the distributor **310**.

The third mixture can flow into the distributor **310** via the inlet **312** and split or be divided among one or more outlets **314**. The third mixture can then flow out the distributor **310** via the outlets **314** and into the containers **320**, as shown in FIG. 4. The third mixture can be introduced to the vacuumed space in the containers **320** between the second end **326** and the plug **328**. In at least one embodiment, the containers **320** can include a fourth reagent (which may be dried or liquid) disposed therein. Thus, the third mixture can be combined with the fourth reagent in the containers **320** to form a fourth mixture therein. The volume of the fourth reagent in each container **320** can range from a low of about zero (0, i.e., dried form), 0.01 mL, about 0.02 mL, about 0.05 mL, about 0.10, about 0.2 mL, about 0.5 mL, or about 1.0 mL to a high of about 2.0 mL, about 3.0 mL, about 4.0 mL, about 5.0 mL, or more. In at least one embodiment, the fourth reagent can be suspended, or bind to the bottom of the container, including oligonucleotide primers, probes, enzymes, buffers, inhibitors, activators in either dried form or liquid form, fluorophore labeled or colorimetric analysis, or the like.

Once the third mixture has been introduced to the containers **320**, the outlets **314** of the distributor **310** can then be withdrawn from the containers **320**. The seals **322** and the plugs **328** can secure the fourth mixture within the containers **320**. A cap (not shown) can also be placed on the first end **322** of the container **320** to add an additional sealing element. For

example, the cap can be threaded onto the first end **322** of the container **320**. The container **320** can then be used for polymerase chain reactions or other processes.

FIGS. 5A-5C depict perspective cross-sectional views of another illustrative embodiment of the preparation assembly **200** in fluid communication with the containers **320** via the distributor assembly **300**, according to one or more embodiments. As shown, the preparation assembly **200** can include a second valve **550** in fluid communication with the first valve **250**. In another embodiment, the preparation assembly **200** can include a second valve **550** in fluid communication with the fourth injector **240** (not shown). One or more injectors (two are shown **510**, **520**) can be coupled to the second valve **550**.

In at least one embodiment, the first and second valves **250**, **550** can be rotated to form a flow path into a fifth reservoir **514** in the fifth injector **510**. Referring to FIG. 5A, the third piston **232** can be pressed, thereby forcing at least a portion of the second mixture through the flow path **584** into the fifth reservoir **514**, forming a third mixture including the second mixture and a fourth reagent. The fourth reagent can include any one or more of the reagents discussed above.

The second valve **550** can then be rotated to provide a flow path **586** as shown in FIG. 5B between the fifth reservoir **514** and the containers **320** (via the distributor assembly **300**). The fifth piston **512** can be pressed, thereby forcing at least a portion of the third mixture through the distributor assembly **300** and into at least one of the containers **320**. As shown in FIG. 5C, the second valve **550** can then be rotated to form a flow path **588** between the sixth reservoir **524** and the containers **320** via the distributor assembly **300**. The sixth piston **522** can then be pressed, thereby forcing at least a portion of a fifth reagent into the containers **320** to mix with the third mixture. The fifth reagent can include any one or more of the reagents discussed above.

In at least one embodiment, prior to transferring the mixture (e.g., from the fifth reservoir **514** and/or the sixth reservoir **524**, or from the fourth reservoir **244**) from the preparation assembly **200** to the containers **320** via the distributor assembly **300**, the distributor **310** can be soaked or rinsed in one or more buffers (such as by placement in a container of buffer solution or by introducing buffer(s) through the inlet **312**). Suitable buffers can include, but are not limited to, deionized water, ribonuclease (RNase) free water, or the like. After soaking in the buffer solution, the mixture can be drawn into the distributor **310** until the mixture approaches the tip of the outlets **314**. At this time, the outlets **314** can be inserted into the containers **320**. By drawing the mixture into the distributor **310** and proximate the tip of the outlets **314**, bubbles may be reduced or prevented in the containers **320**.

FIG. 6 depicts a perspective view of a plurality of preparation assemblies **200**, **600**, **700**, **800** in fluid communication with the containers **320** via the distributor assembly **300**, according to one or more embodiments described. A plurality of preparation assemblies (four are shown **200**, **600**, **700**, **800**) can be in fluid communication with the containers **320** via the distributor **310**. As shown, tubes **248**, **648**, **748**, **848** may be coupled respectively to the corresponding fourth injector **240**, **640**, **740**, **840** of each preparation assembly. Tubes **248**, **648**, **748**, **848** may provide a path of fluid communication from each respective fourth reservoir **244**, **644**, **744**, **844** to the distributor assembly **300**. Each tube **248**, **648**, **748**, **848** can be coupled to the same distributor assembly **300**, or each tube **248**, **648**, **748**, **848** can be coupled to a different distributor assembly (not shown).

Certain embodiments and features have been described using a set of numerical upper limits and a set of numerical

lower limits. It should be appreciated that ranges from any lower limit to any upper limit are contemplated unless otherwise indicated. Certain lower limits, upper limits, and ranges appear in one or more claims below. All numerical values are “about” or “approximately” the indicated value, and take into account experimental error and variations that would be expected by a person having ordinary skill in the art. The reagents listed are mainly for real-time PCR or RT-PCR. For other applications, such as enzymatic assay, luminal assay, ELISA, the reagents may vary.

Various terms have been defined above. To the extent a term used in a claim is not defined above, it should be given the broadest definition persons in the pertinent art have given that term as reflected in at least one printed publication or issued patent. Furthermore, all patents, test procedures, and other documents cited in this application are fully incorporated by reference to the extent such disclosure is not inconsistent with this application and for all jurisdictions in which such incorporation is permitted.

Although only a few exemplary embodiments have been described in detail above, those skilled in the art will readily appreciate that many modifications are possible in the exemplary embodiments without materially departing from the novel teachings and advantages of this invention. Accordingly, all such modifications are intended to be included within the scope of this invention as defined in the following claims. In the claims, means-plus-function and step-plus-function clauses are intended to cover the structures or acts described herein as performing the recited function and not only structural equivalents, but also equivalent structures. Thus, although a nail and a screw may not be structural equivalents in that a nail employs a cylindrical surface to secure parts together, whereas a screw employs a helical surface, in the environment of fastening parts, a nail and a screw may be equivalent structures.

What is claimed is:

1. A preparation system, comprising:

a first injector having a first reservoir disposed therein;

a second injector having a second reservoir disposed therein;

a valve disposed between the first and second reservoirs such that a path of fluid communication exists between the first and second reservoirs through the valve;

a membrane disposed between the first reservoir and the valve such that the path of fluid communication extends through the membrane;

a distributor having an inlet coupled to the second injector and in fluid communication with the second reservoir; and

one or more containers coupled to an outlet of the distributor, wherein a reagent when disposed in the second reservoir is capable of flowing from the second reservoir, through the distributor, and into the one or more containers.

2. The preparation system of claim 1, wherein the second injector comprises a piston having a bore formed there-through, and wherein the second reservoir is in fluid communication with the inlet of the distributor through the bore.

3. The preparation system of claim 1, wherein the membrane comprises nucleic acids bound thereto, and wherein when a reagent disposed in the second reservoir contacts the membrane, at least a portion of the nucleic acids are adapted to release from the membrane and become disposed within the reagent.

4. The preparation system of claim 1, further comprising a reagent disposed in the second reservoir.

5. The preparation system of claim 4, further comprising a third injector coupled to the valve and having a third reservoir disposed therein, wherein the third reservoir comprises a polymerase chain reaction master mix disposed therein.

6. The preparation system of claim 1, wherein each of the one or more containers comprises a plug disposed therein, wherein the plug is adapted to sealingly engage an inner wall of the container.

7. The preparation system of claim 1, wherein the one or more containers are pre-vacuumed by moving a plug disposed therein.

8. A method for preparation of a biological sample for analysis using the preparation system of claim 1, wherein the method comprises contacting the biological sample with a reagent adapted to flow from the second reservoir, through the valve and the membrane, and into the first reservoir.

9. The method of claim 5, wherein the reagent comprises between about 0.1 mL and about 3.0 mL of water.

10. The method of claim 8, wherein the reagent comprises between about 0.1 mL and about 3.0 mL of a polymerase chain reaction master mix.

11. The preparation system of claim 1, wherein the second injector comprises a piston disposed at least partially therein, and wherein a bore extends through the piston such that the bore is in fluid communication with the second reservoir.

12. The preparation system of claim 1, further comprising a spectrophotometer in communication with an interface on the first injector, the second injector, or both.

13. The preparation system of claim 1, wherein at least one of the first and second injectors is threadably engaged with the valve.

14. A method of releasing nucleic acids from a membrane, comprising:

providing a self-contained preparation system comprising a first reservoir, a second reservoir, a membrane for collecting nucleic acids disposed between the first reservoir and the second reservoir, and a valve disposed between the first reservoir and the second reservoir and the membrane such that a path of fluid communication exists between the first and second reservoirs through the valve;

flowing a first reagent through a first flow path between the first reservoir and the second reservoir, wherein the first flow path extends through the valve and the membrane, and wherein nucleic acids are released from the membrane when contacted with a first reagent to form a first mixture comprising the first reagent and the released nucleic acids;

positioning the valve to form a second flow path between the second reservoir and a third reservoir through the valve; and

flowing at least a portion of the first mixture through the second flow path and into the third reservoir containing a second reagent to form a second mixture.

15. The method of claim 14, wherein at least one of the first and second reagents comprises water.

16. The method of claim 14, further comprising measuring a concentration of the nucleic acids disposed within the first mixture.

17. The method of claim 14, further comprising flowing the second mixture through a distributor and into one or more containers for further analysis.